

# *Non-Destructive Digital Imaging of Poplar Root Systems.*

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# Why are roots important?

- Water uptake and movement.
- Mineral uptake and movement.
- Support
- Storage
- Mutualism: Interaction with organisms
  - Mycorrhizae association
  - Root nodulation bacteria
- Parasites
  - Nematodes and others
- Disease Resistance





# Root Systems are Difficult to Analyze

## Common Methods:

- Invasive
- Destructive.
- Time consuming
- Provide limited information

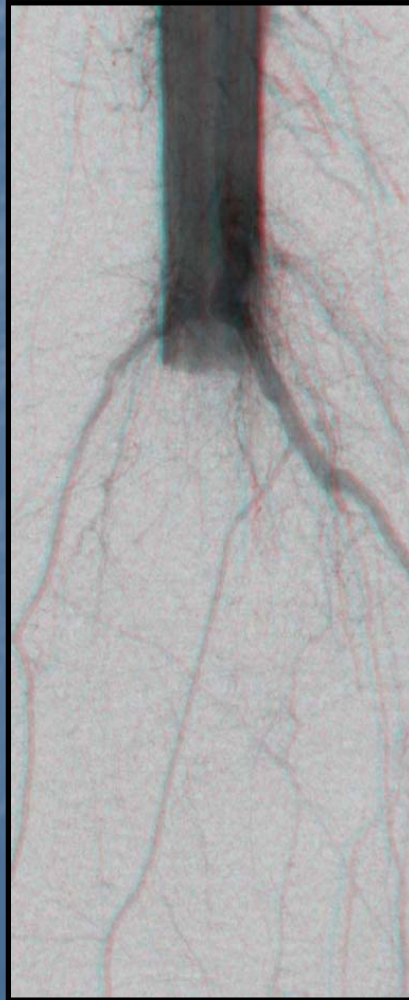


# RootViz FS: A New Way to Study Roots

Based on low energy x-ray imaging

## Features:

- Non-destructive
- Non-invasive
- High resolution
- Versatile
- Quantitative
- High throughput

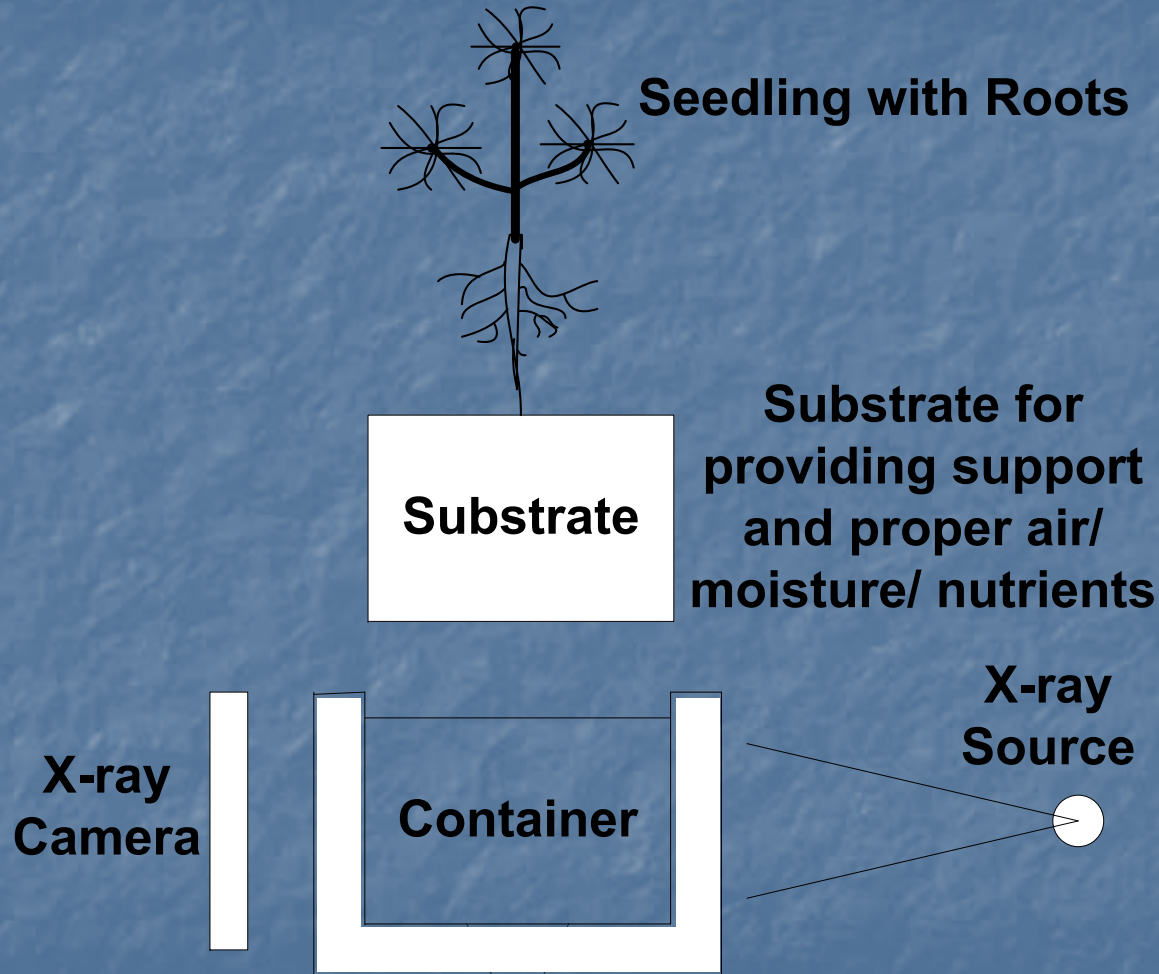


## Capacities:

- 0.6 m rooting depth
- 2.1 m plant height
- Up to 25 plants per hour throughput
- Seed germination
- Greenhouse stage plants



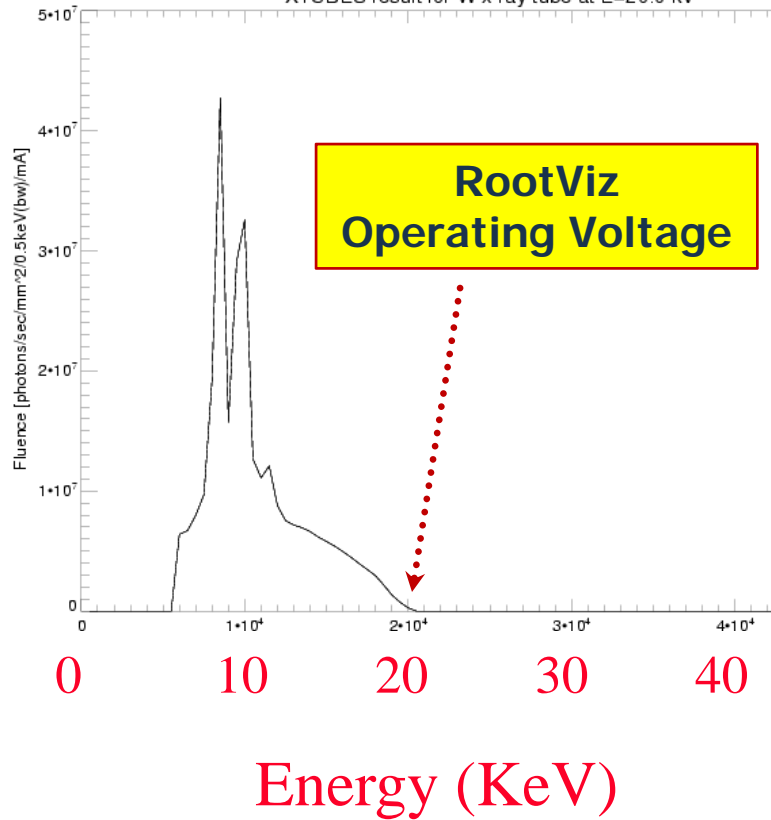
# Key Components of Concept



**Container for substrate and seedling.  
Allows for plant storage and transport.**

# Low Energy X-ray System

XTUBES result for W x-ray tube at E=20.0 kV



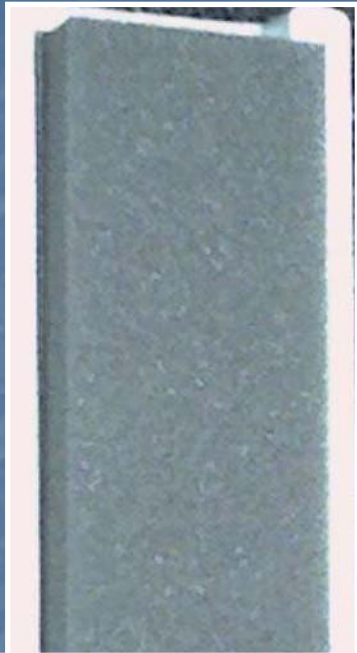
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In collaboration with:  
ArborGen, Oak Ridge National Laboratory, University of Tennessee



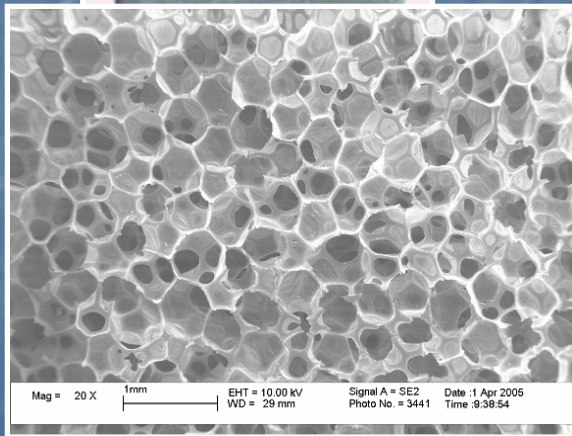
# Engineered Rooting Substrates



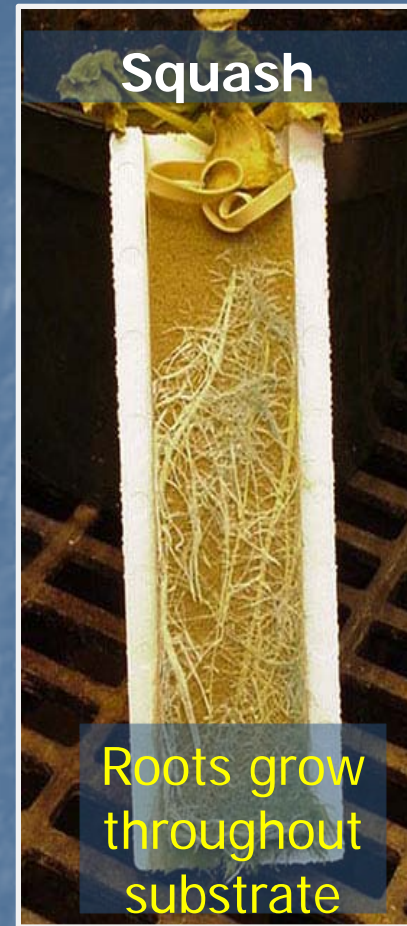
Like agar or special soil mixtures, the PSC engineered rooting substrate provides a specific set of conditions for each experiment.

Unlike soil mixtures, PSC engineered rooting substrate provides more precise control.

- Consistent conditions over the entire rooting environment.
- Lower environmental variation among plants and between experiments.
- Rapid changes to the rooting environment can be easily introduced.



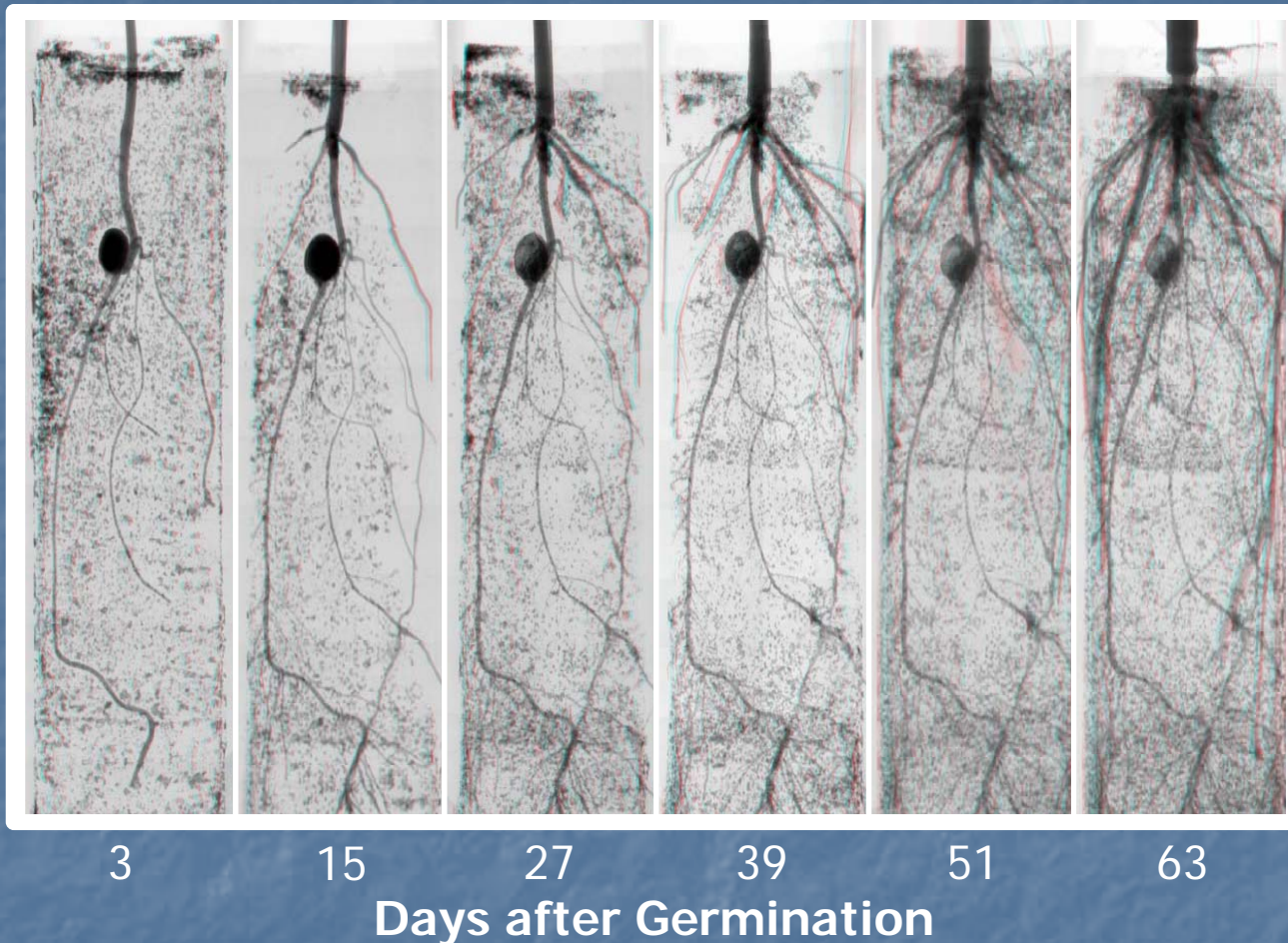
# Plant Growth in PSC Substrates



- An engineered rooting substrate allows normal root growth and enables x-ray analysis.
- A wide range of species can be analyzed.



# Powerful Developmental Studies

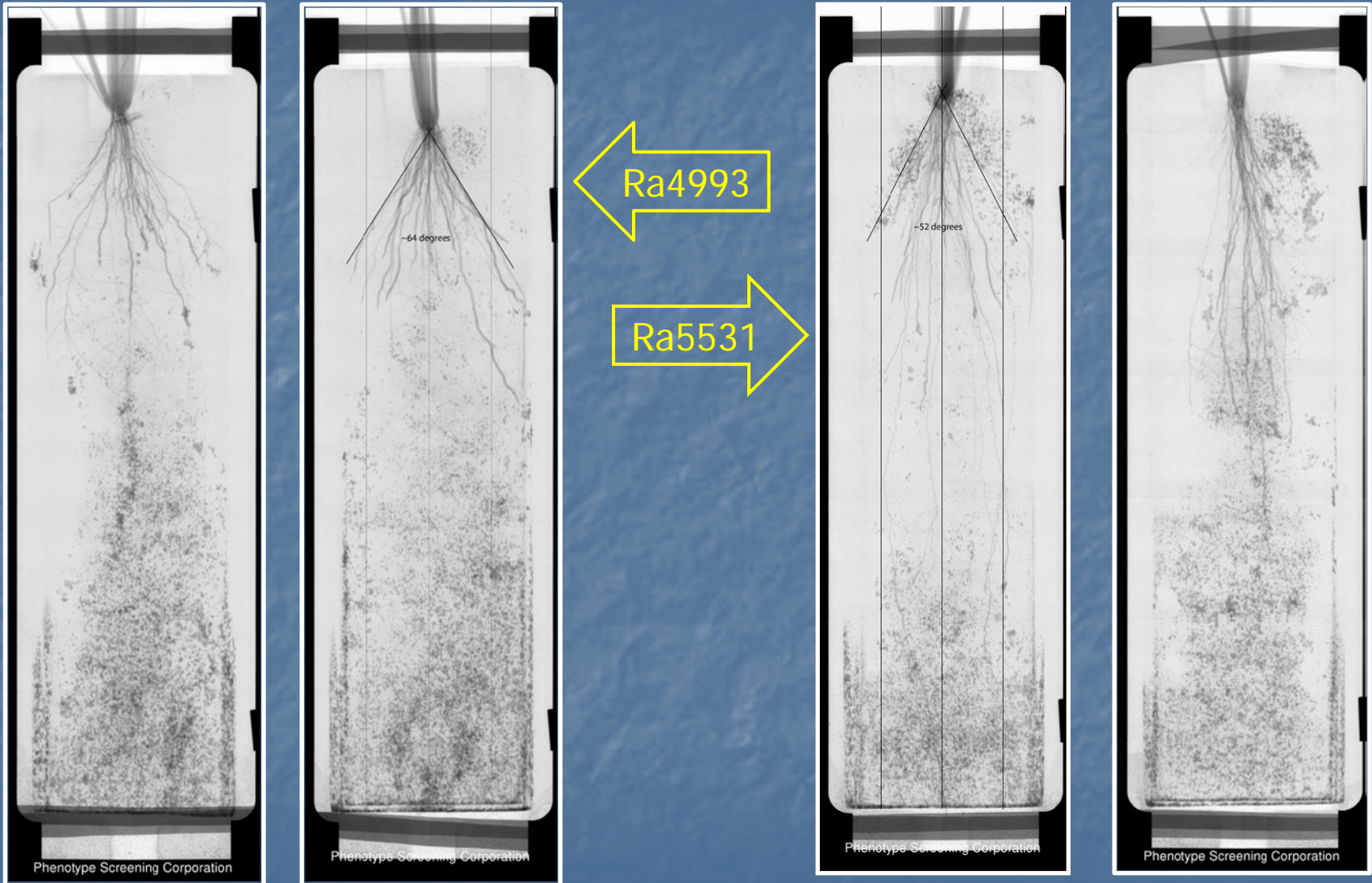


A single maize plant was imaged at 12 day intervals showing the development of the root system.

Data generated in collaboration with:

Dr Edward Buckler, USDA-ARS, Cornell University, Ithaca, NY; Dr Philip Benfey, Dept of Biology, Duke University, Durham, NC

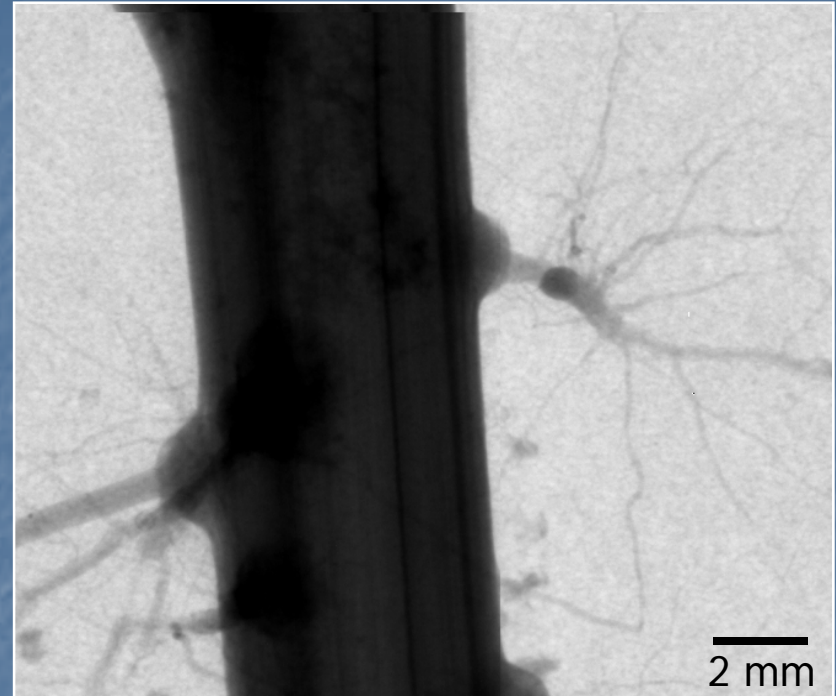
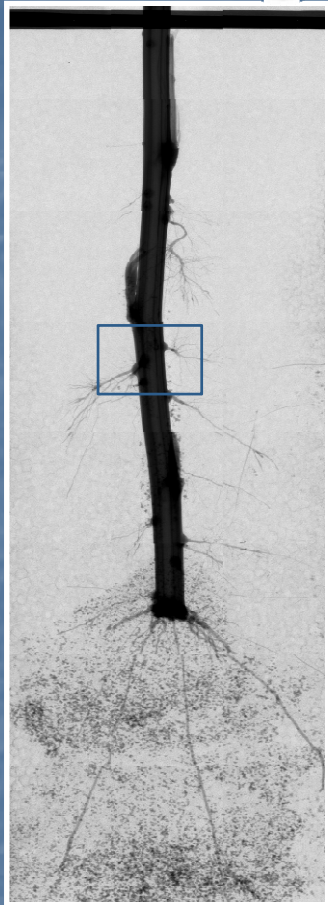
# Rice: Variety Attributes



Data generated in collaboration with:  
Dr Susan McCouch, Dept of Plant Biology, Cornell University, Ithaca, NY  
Dr Philip Benfey, Dept of Biology, Duke University, Durham, NC



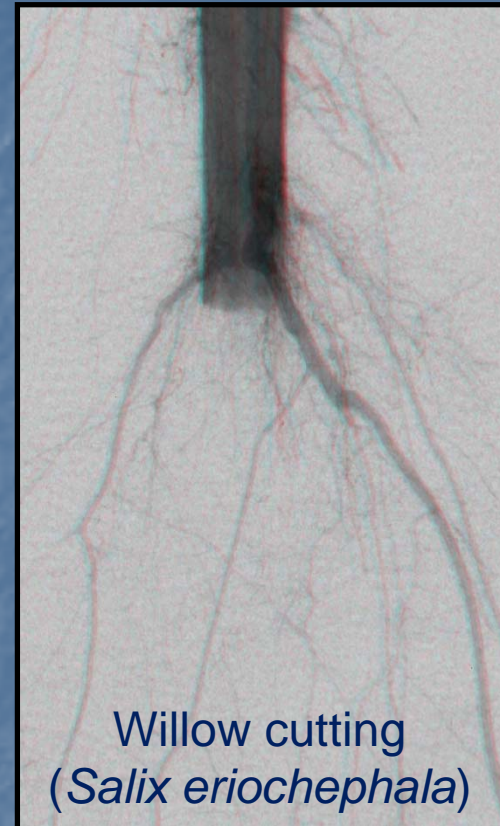
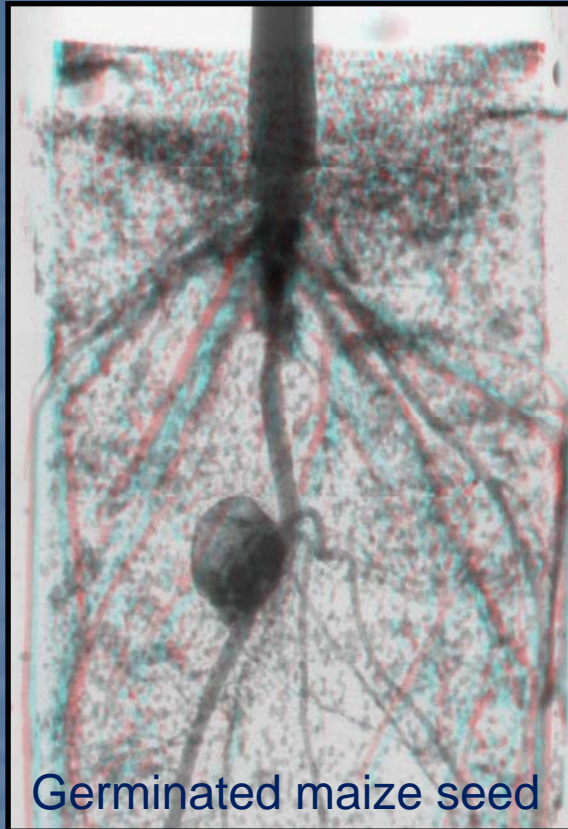
# High Resolution X-Ray Images



## Detailed Analysis in 2D (Mono)

The x-ray images are taken at 100 micron resolution. Zooming in on an area shows a high level of detail. The close-up shows root nodes on the front or back surface of the root as well as the smaller roots crossing the original poplar cutting.

# High Resolution Stereo Images



RootViz FS is capable of taking high resolution stereo images. This is achieved by taking two images of the root system with a slight rotation of the plant in the x-ray chamber.

*Note: Stereo viewing requires red/cyan glasses.*



# RootViz Summary

- A potentially useful tool for root studies
  - Morphology
  - Development
  - Screening
  
- But...

# It's not soil...

- How does growth in engineered substrate compare to soil?
- Are the results accurate?
- Are the results meaningful?



# Collaboration with USDA Forest Service

- Compare growth of poplar clones between engineered substrates and soil.
- Compare traditional root analytical methods with x-ray analysis.
- Suggest modifications to foam substrates.
- Suggest improvements to x-ray characterization system.



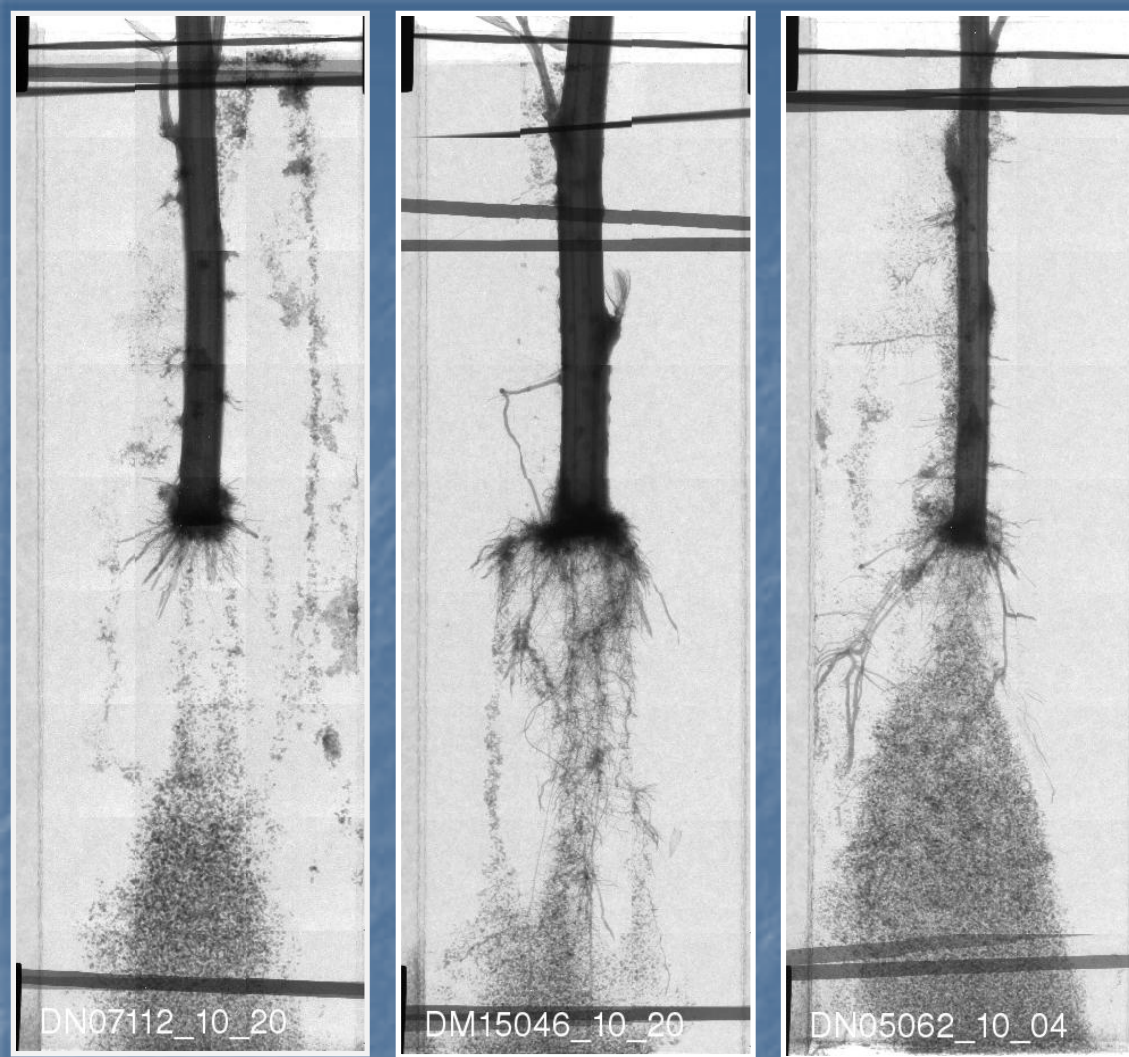
# Collaborative Project with US Forest Service to Study Poplar Rooting

- 3 poplar clones: 4 replicates per clone
  - DN70 *P. deltoides* x *P. nigra*
  - NC14104 *P. deltoides* x *P. maximowiczii*
  - NM6 *P. nigra* x *P. maximowiczii*
- 4 Rooting Media
  - Sand
  - Peat/vermiculite mix
  - LWR engineered rooting matrix
  - MWR engineered rooting matrix





# Root Development



Data generated in collaboration with:  
Dr Alex Friend, Northern Research Station  
US Forest Service, Houghton, MI

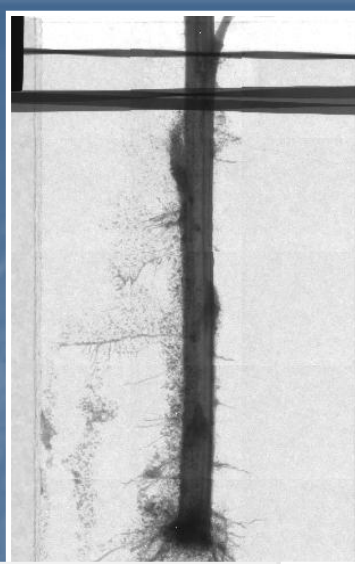
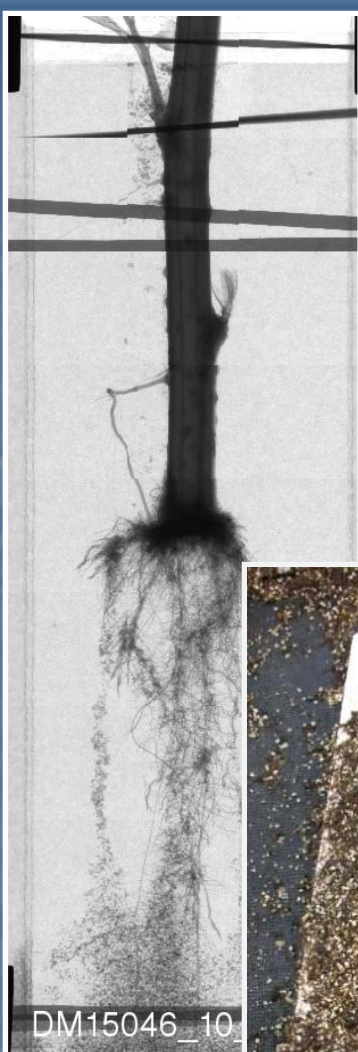
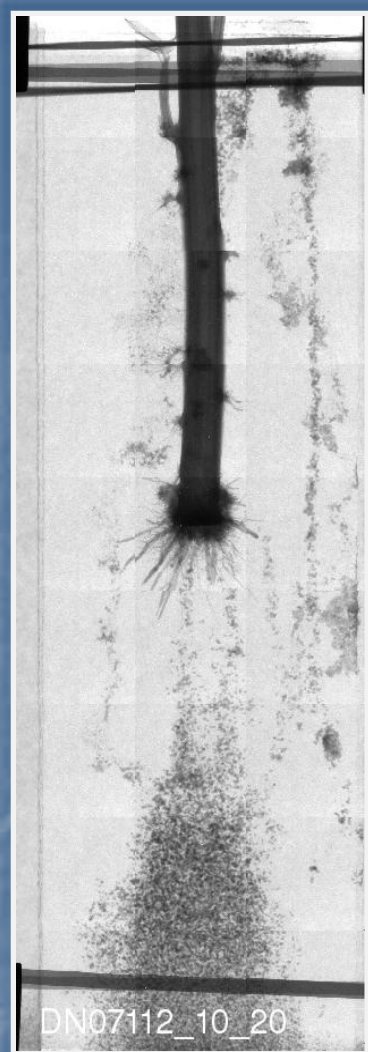


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# Root Development





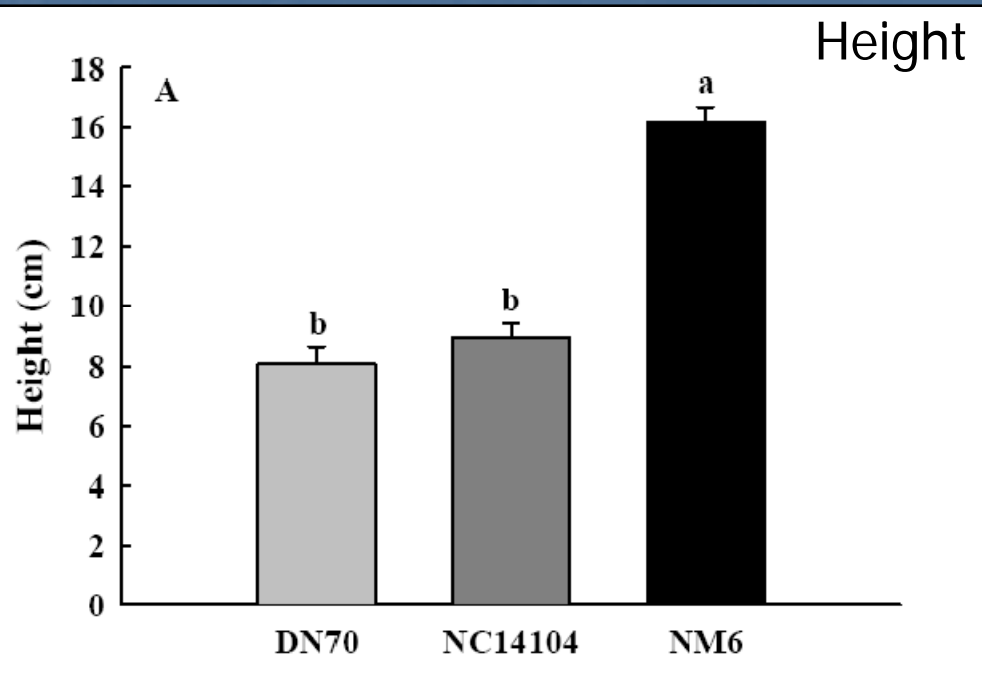
# Poplar Traits Analyzed

Trait Analyzed      Substrate Clone      Substrate Cutting  
X Clone      Dry Mass

Height (cm)	0.1305	<0.0001	0.2977	<b>0.0004</b>
Leaf area (cm <sup>2</sup> )	<b>0.0448</b>	<0.0001	0.5146	<b>0.0201</b>
Leaf dry mass (mg)	<b>0.0294</b>	<0.0001	0.4907	<b>0.0034</b>
Stem dry mass (mg)	<b>0.0252</b>	<0.0001	0.8160	<0.0001
Aboveground dry mass (mg)	<b>0.0266</b>	<0.0001	0.5343	<b>0.0012</b>
Number of primary roots	<b>0.0014</b>	<0.0001	0.8716	0.5506
Number of secondary roots	<0.0001	<b>0.0050</b>	0.6711	<b>0.0091</b>
Number of tertiary roots	<b>0.0002</b>	<b>0.0059</b>	0.0655	0.3052
Length of primary roots (cm)	<0.0001	<0.0001	0.2279	0.3040
Length of secondary + tertiary roots (cm)	<b>0.0002</b>	<b>0.0257</b>	0.3663	0.1510
Dry mass of primary roots (mg)	<b>0.0016</b>	<b>0.0007</b>	0.8766	0.5735
Dry mass of secondary + tertiary roots (mg)	<b>0.0003</b>	<b>0.0227</b>	0.3829	0.5783
Belowground dry mass (mg)	<0.0001	<b>0.0010</b>	0.6624	0.9225
Total tree dry mass (mg)	<b>0.0165</b>	<0.0001	0.5540	<b>0.0022</b>
Root mass fraction	<0.0001	<b>0.0022</b>	0.7284	<b>0.0309</b>
Number of secondary roots per primary root	0.1265	0.1784	0.5822	0.2486
Primary root length per dry mass (cm mg <sup>-1</sup> )	<0.0001	<b>0.0027</b>	0.8606	<b>0.0042</b>
Secondary + tertiary root length per dry mass (cm mg <sup>-1</sup> )	<0.0001	0.3147	0.1837	0.1155



# Clonal Differences



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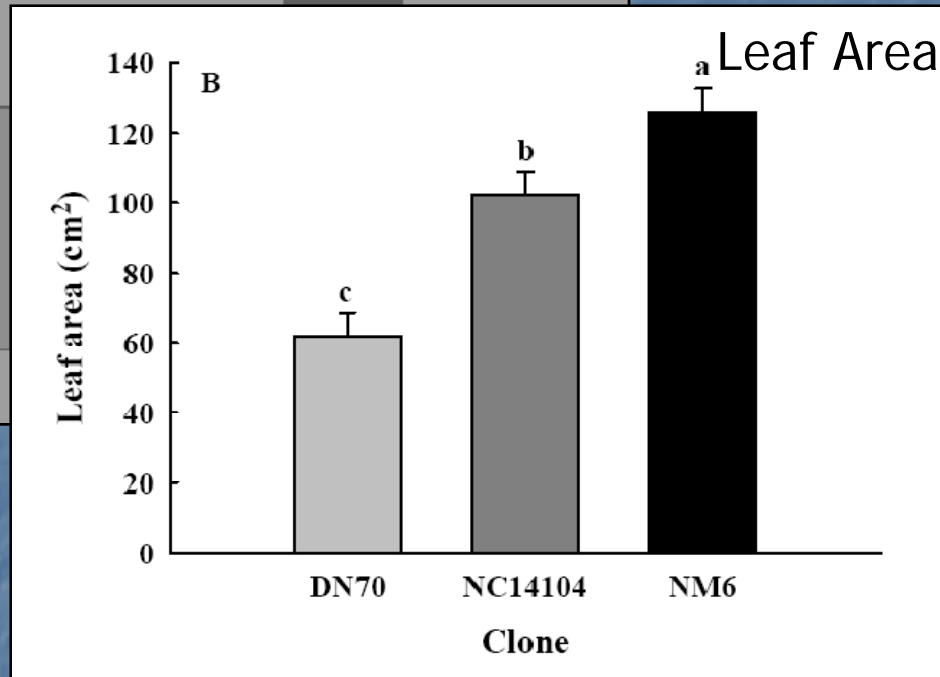
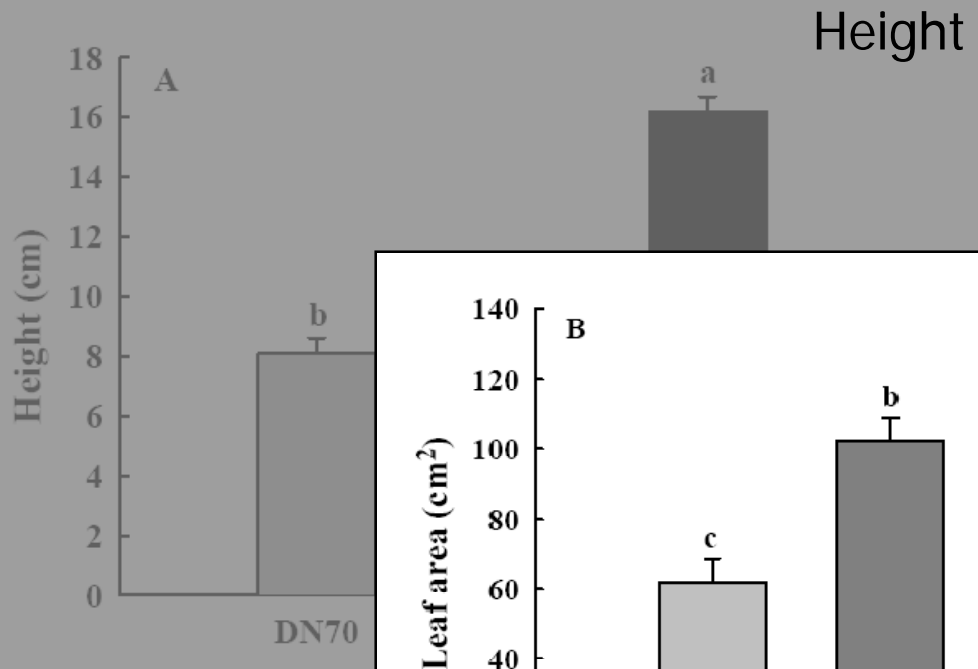


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# Clonal Differences



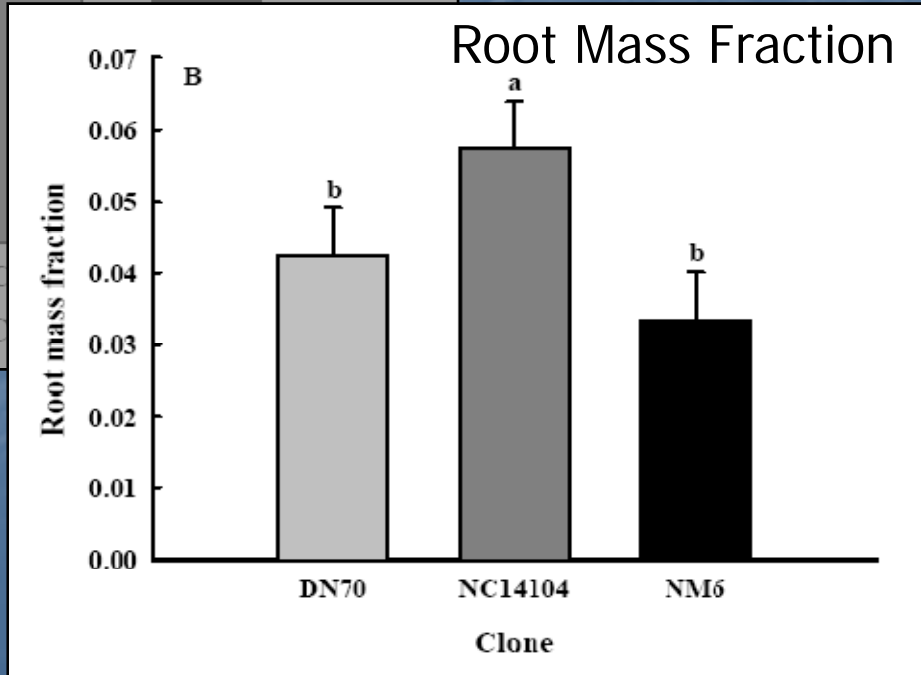
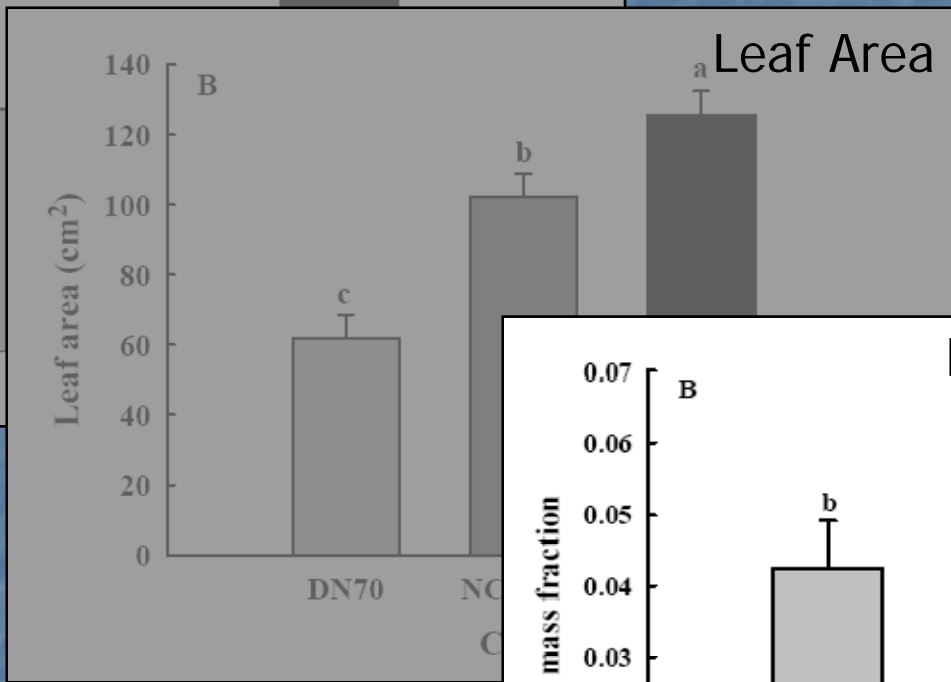
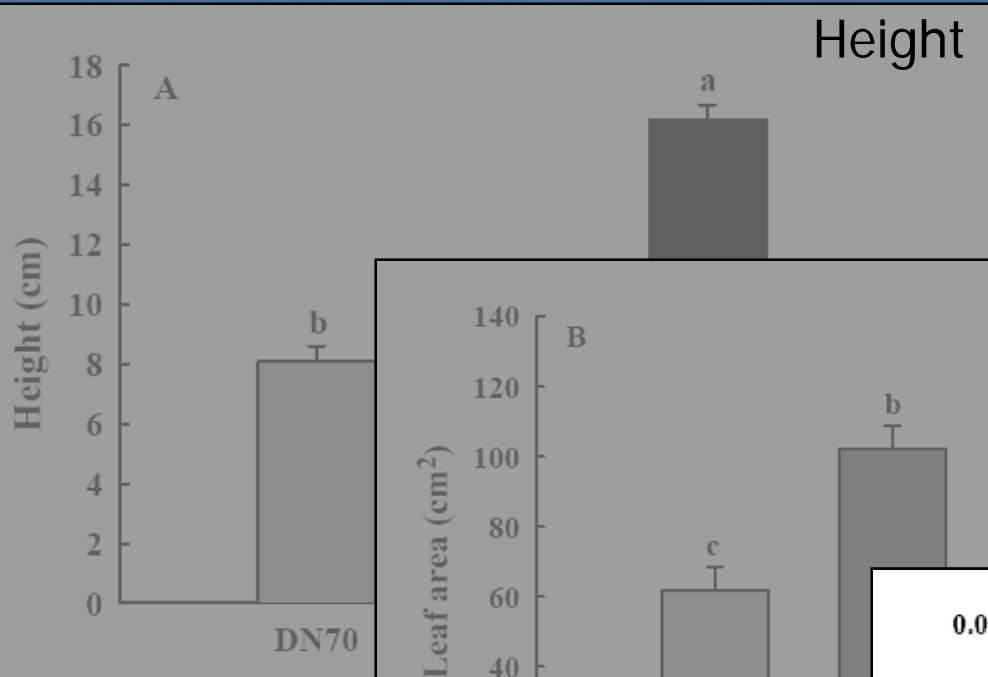
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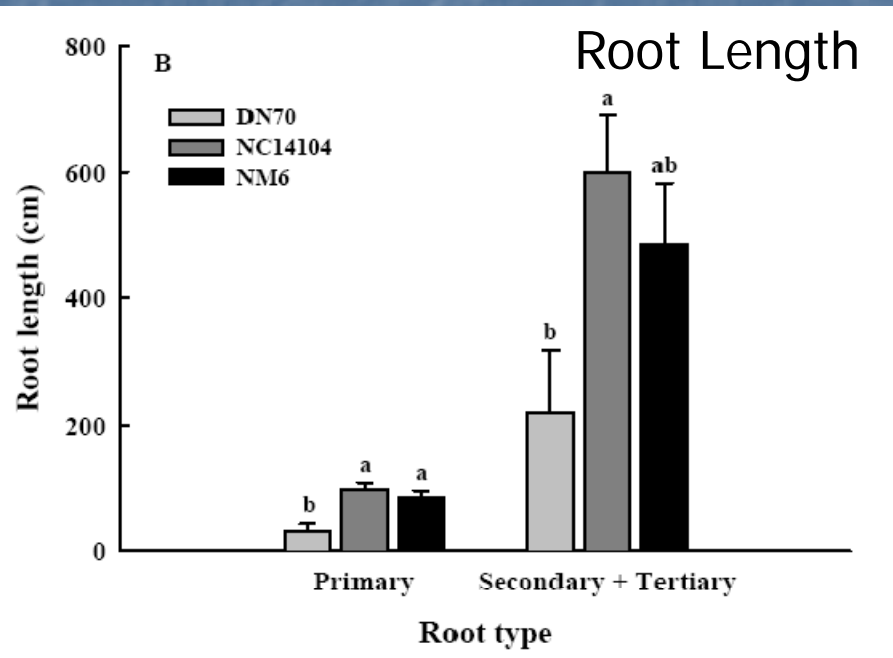


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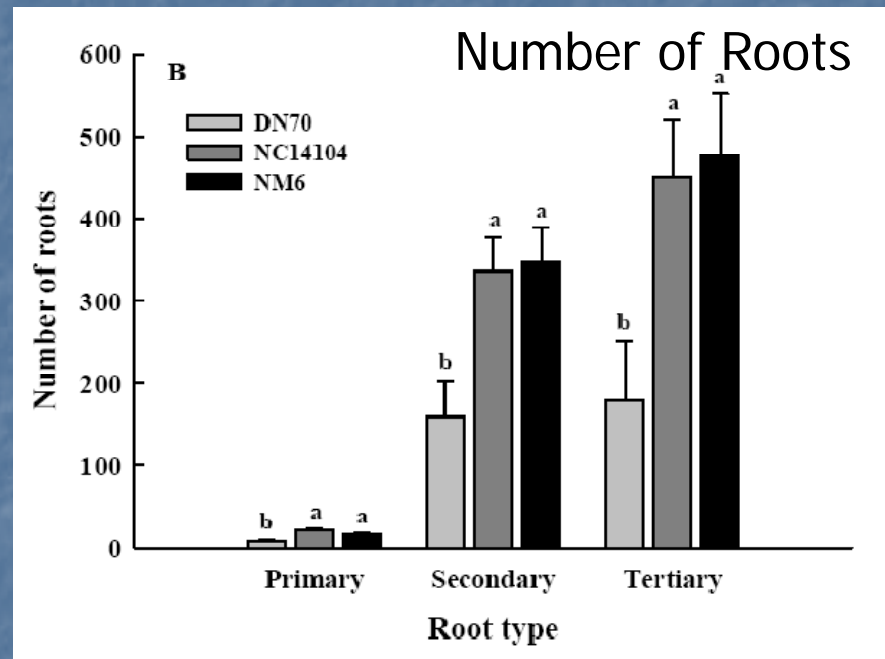
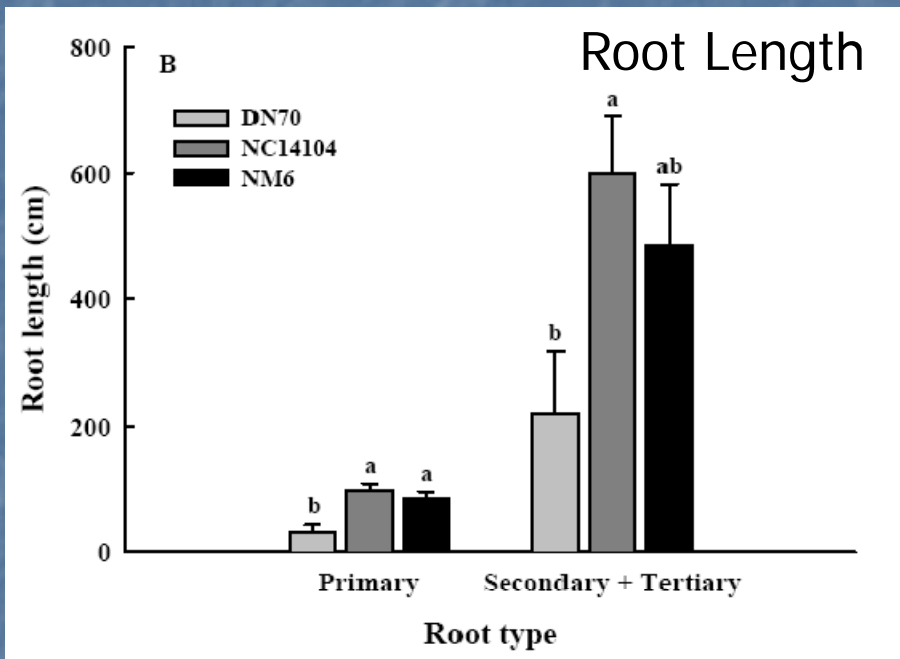




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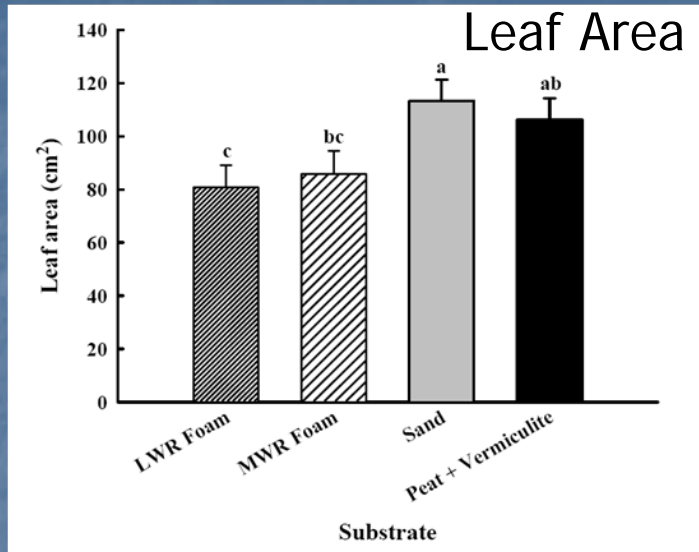


# Clonal Differences





# Substrate Effects



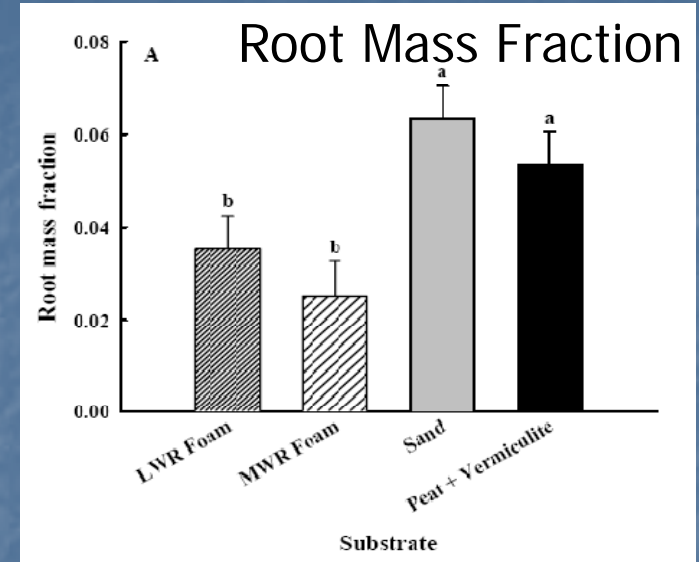
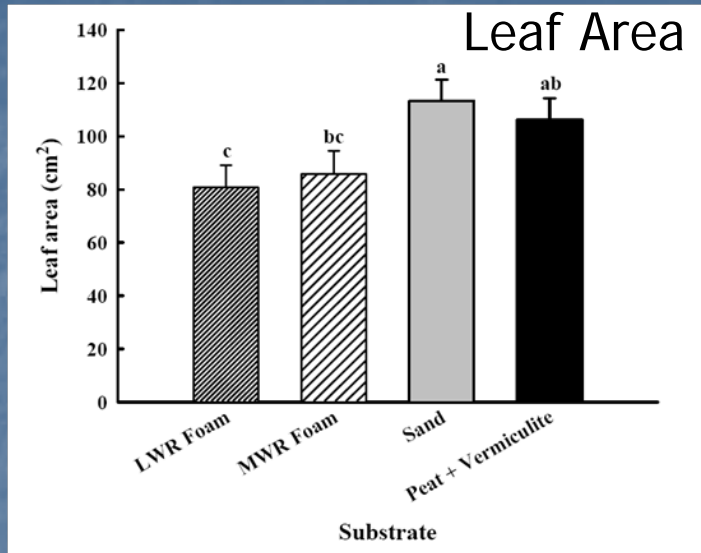
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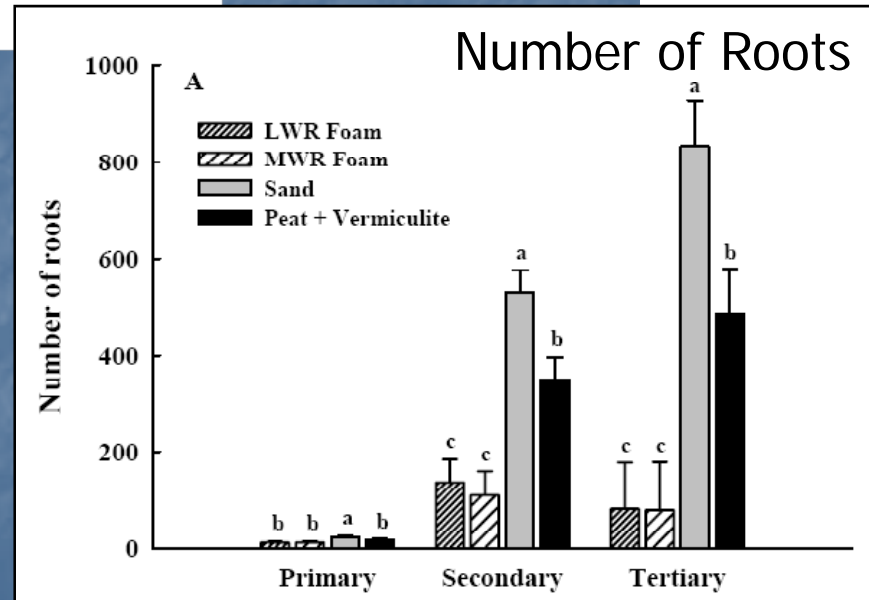
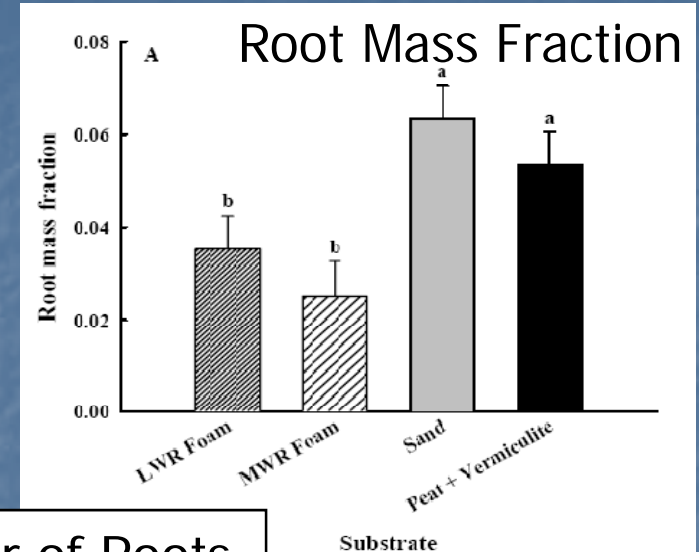
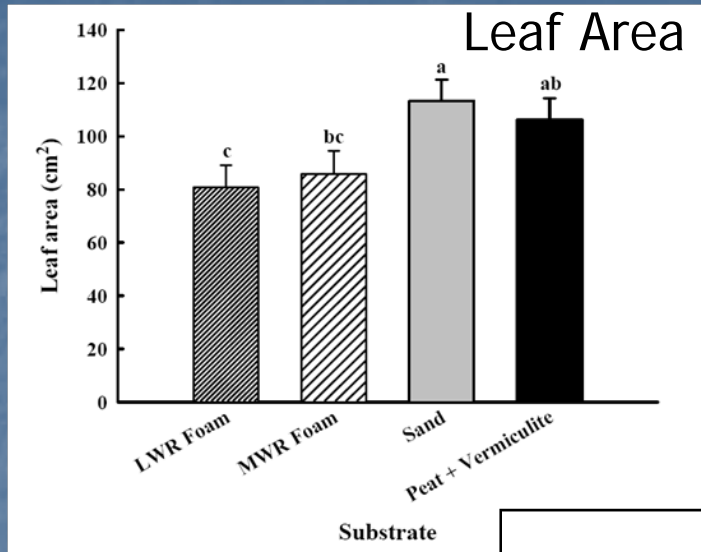


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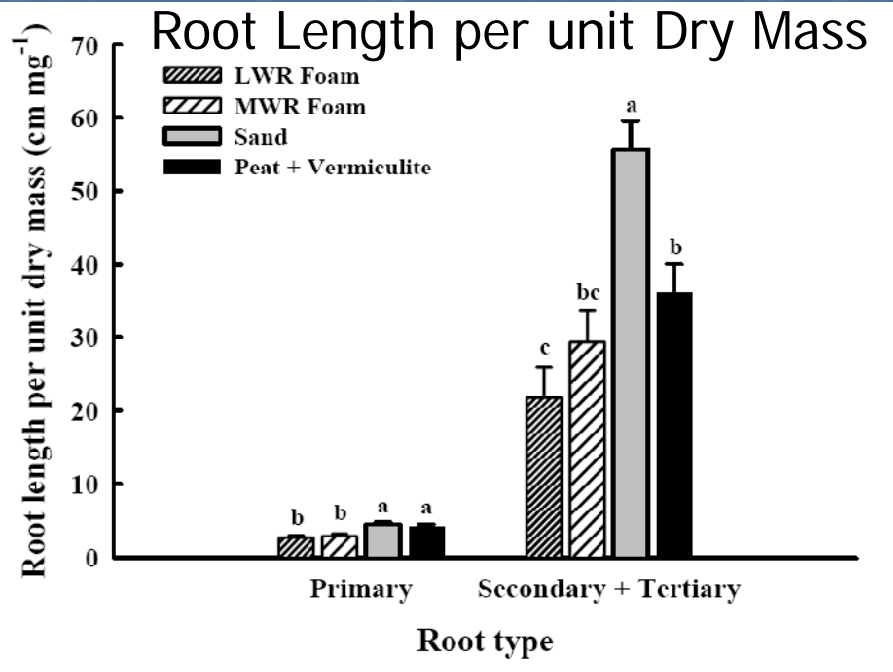




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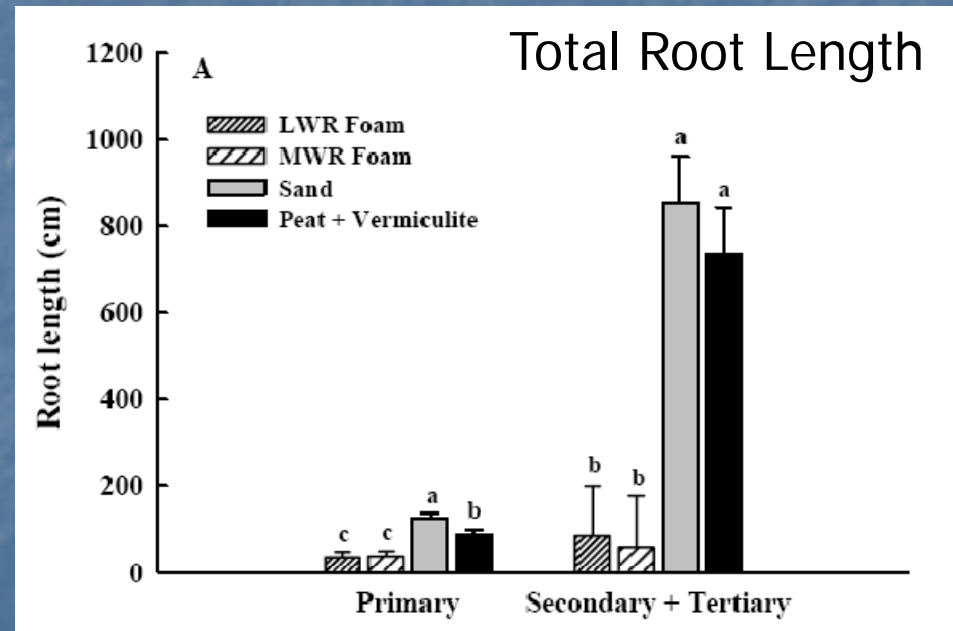
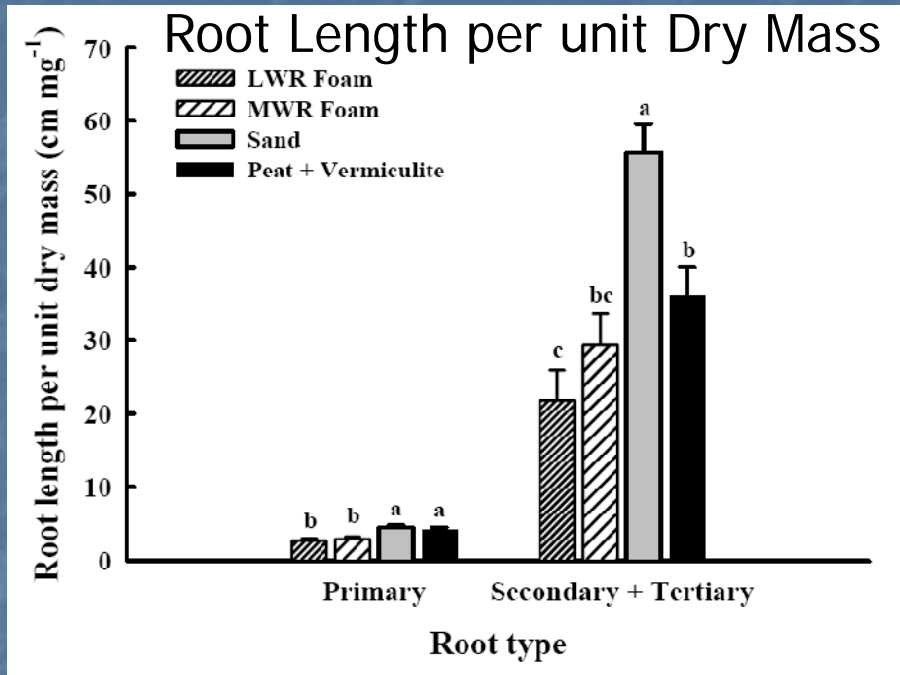


# Substrate Effects





# Substrate Effects



# Clonal Ranking by Substrate

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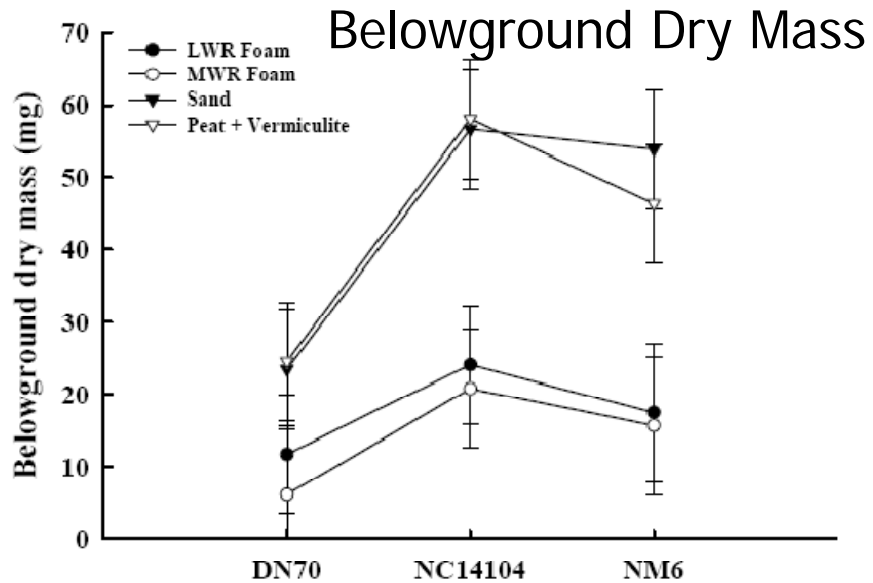


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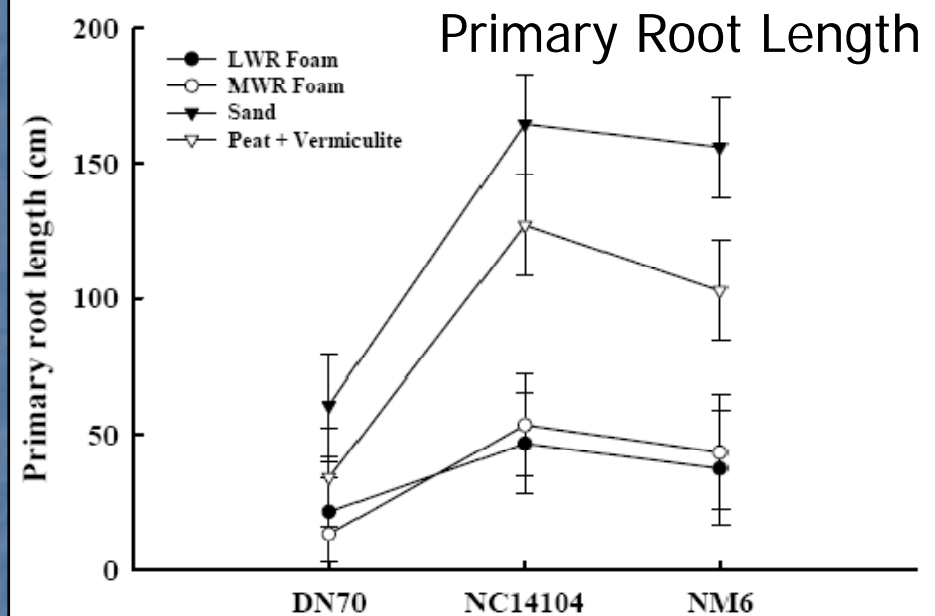
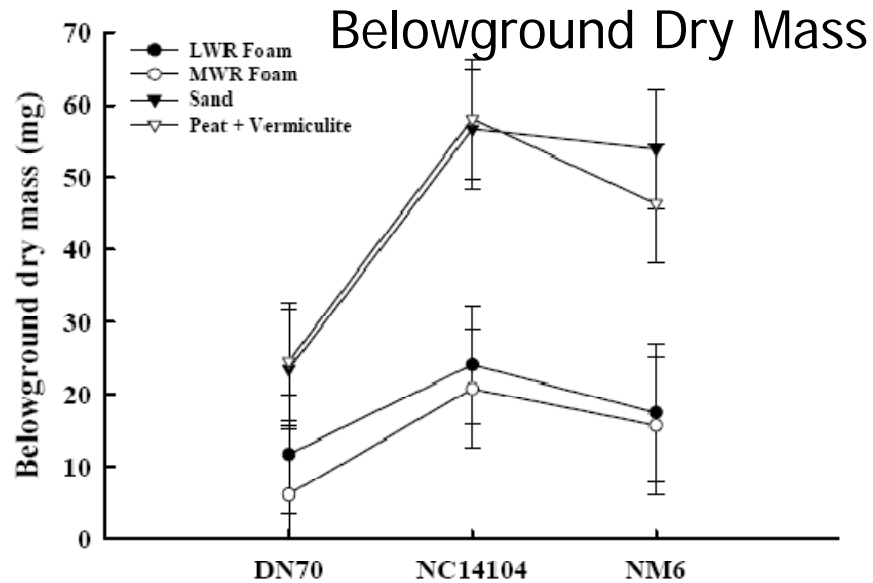




# Clonal Ranking by Substrate



# Clonal Ranking by Substrate



Clonal Rankings Between Engineered Substrates and Soils are Generally Consistent



# Conclusions from Poplar Studies

- Poplars will root and grow, from cuttings, in our engineered substrate materials.
- Root morphology differences were observed between plants grown in "soil" and those grown in engineered substrate.
- Clonal rankings in most root metrics were preserved across substrates.



# Next Step for Poplar Studies

- Experiment is being repeated.
- Addition of nutrients to growth in engineered substrate.
- Results in engineered substrate will be compared to same clones in 1 and 2 year field tests.
- Test ability to predict wind firmness in plantation-grown poplar.
- New substrates will be tested.





# Other Interesting Results...

- Growth dynamics can be measured.
- Compare daytime vs. nighttime growth.
- Detailed morphology studies.





# Distinct Morphotypes of Roots

Naked Pioneer

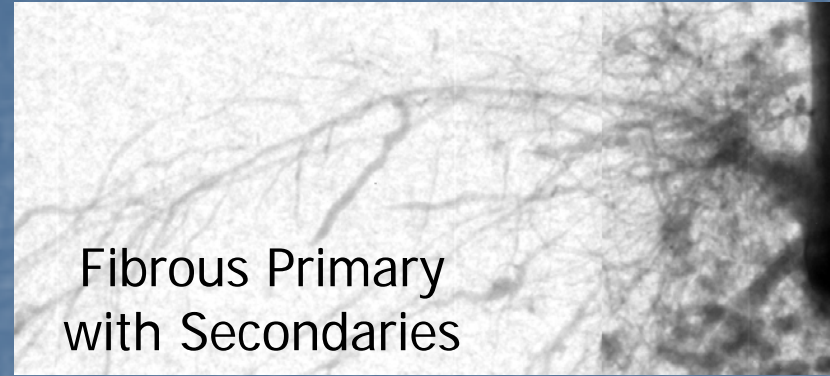


# Distinct Morphotypes of Roots

Naked Pioneer



Fibrous Primary  
with Secondaries

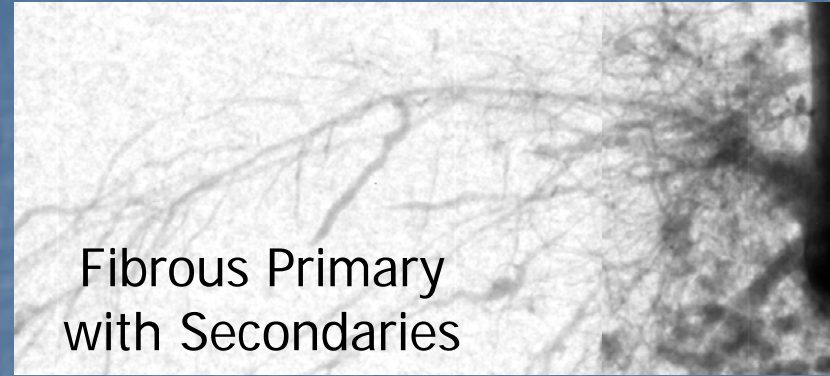


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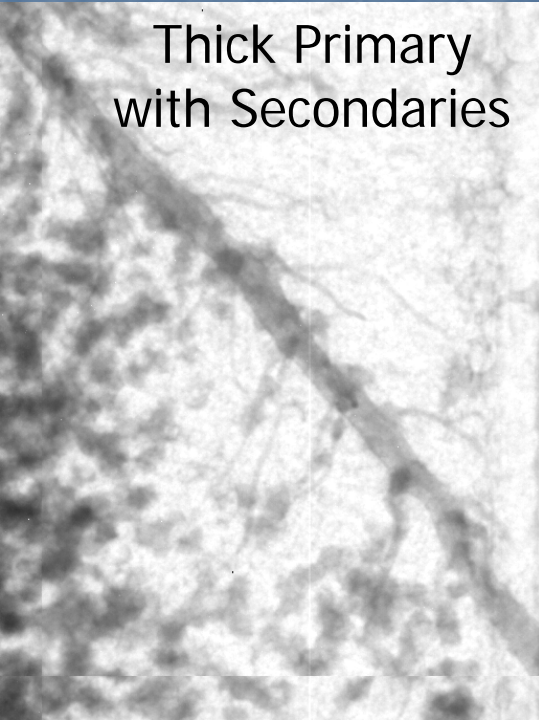
Naked Pioneer



Fibrous Primary  
with Secondaries



Thick Primary  
with Secondaries



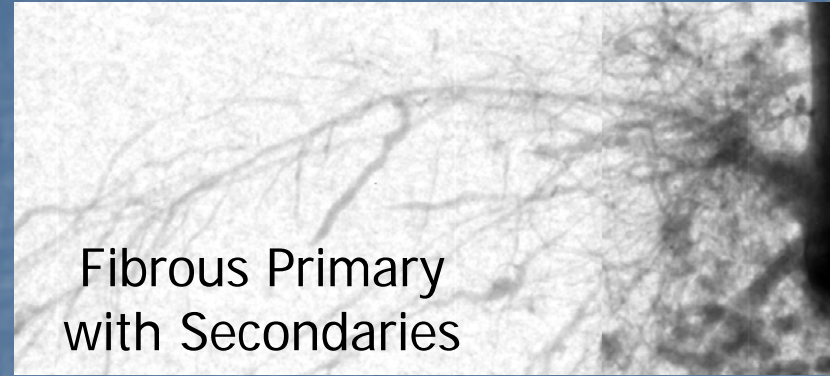


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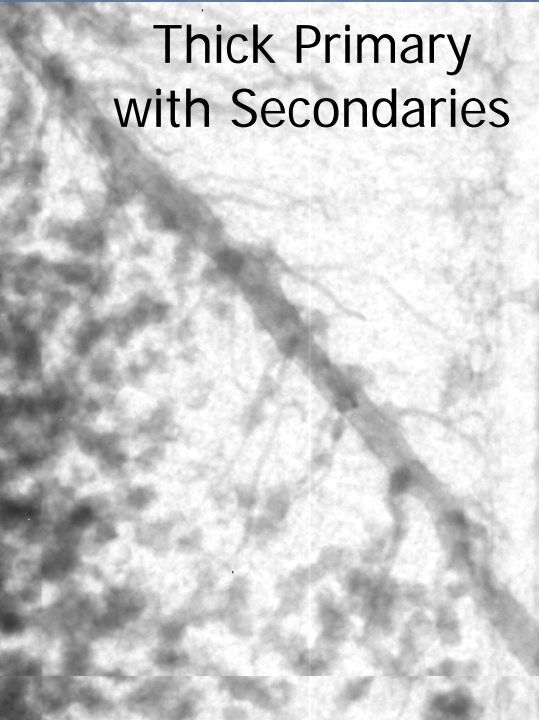
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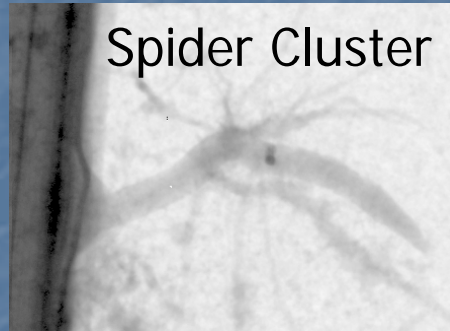
Fibrous Primary  
with Secondaries



Thick Primary  
with Secondaries



Spider Cluster

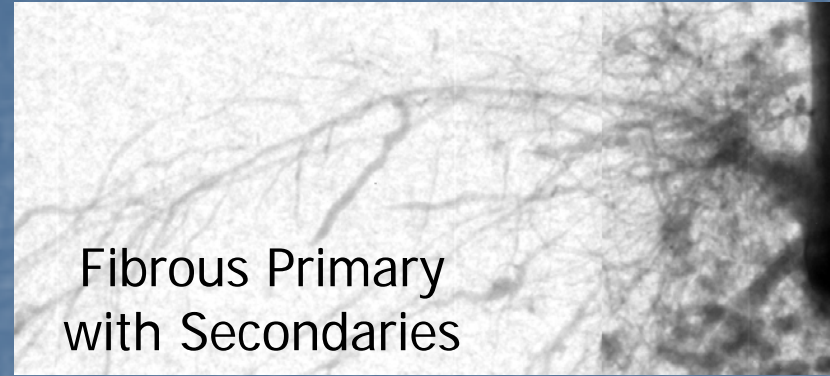


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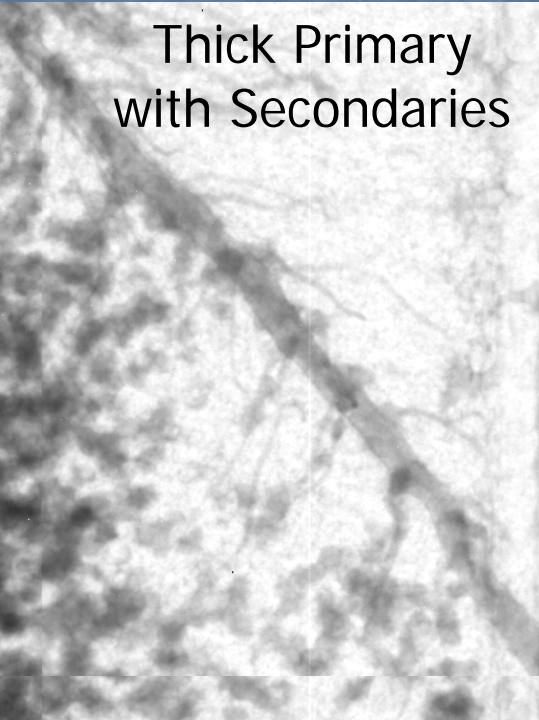
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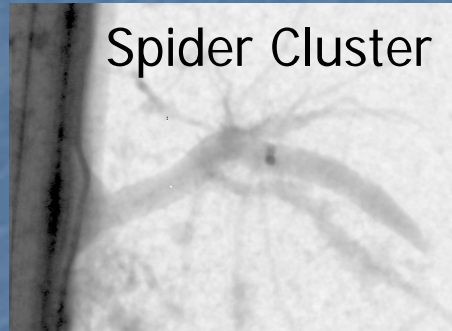
Fibrous Primary  
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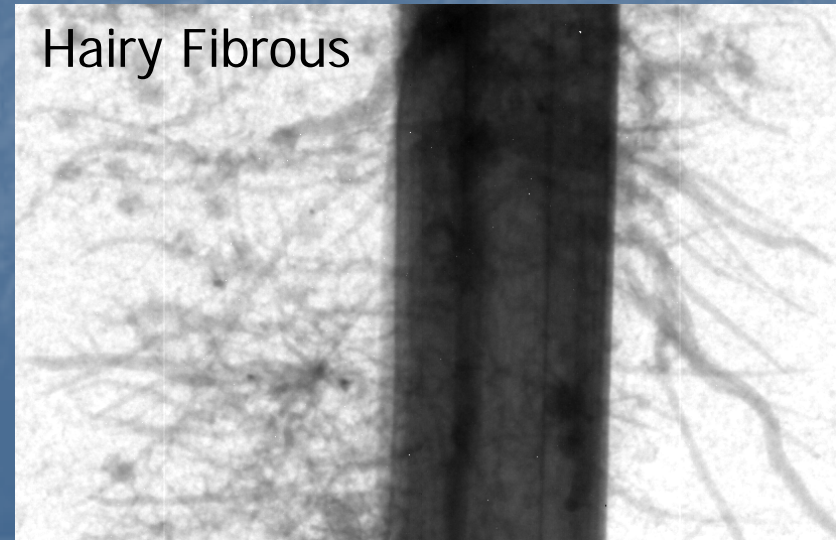
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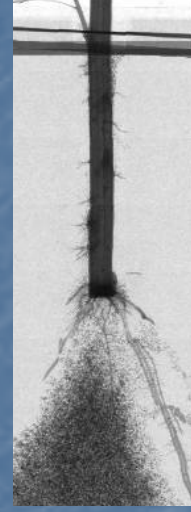
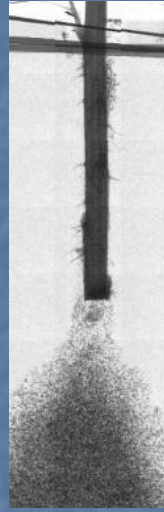


Hairy Fibrous





# Root Growth Delayed after Planting



Days after planting

4

8

12

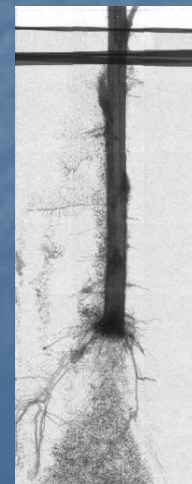
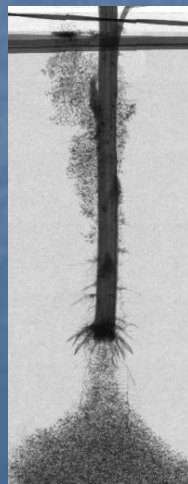
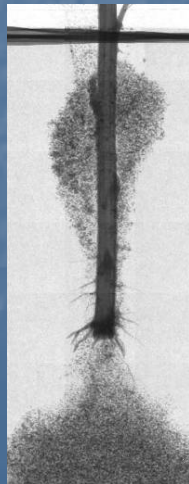
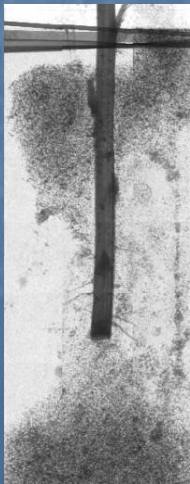
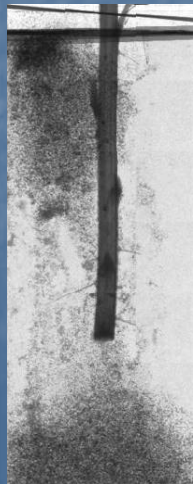
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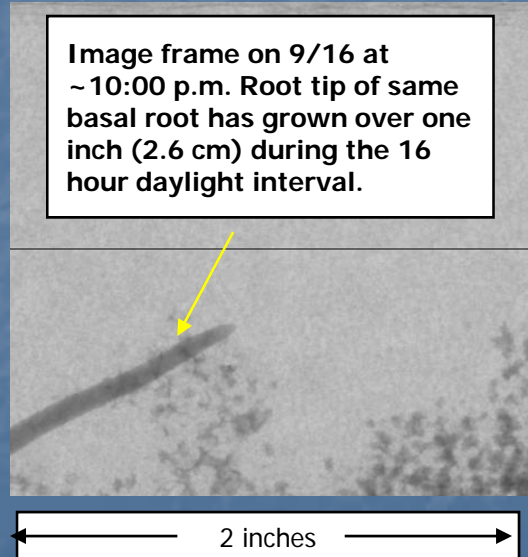
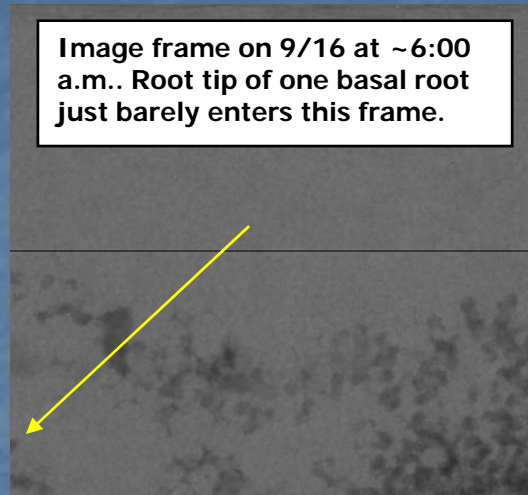
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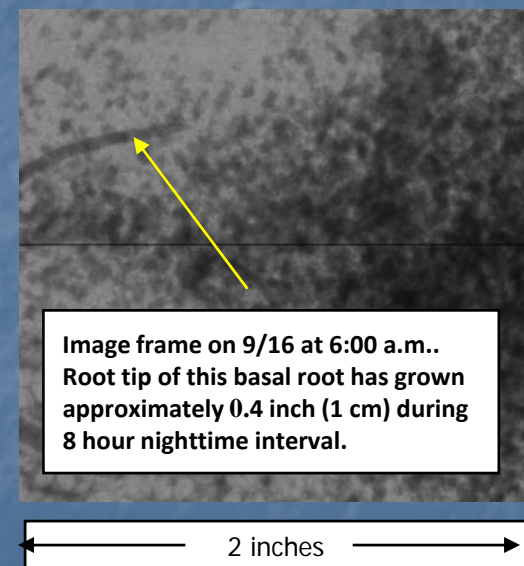
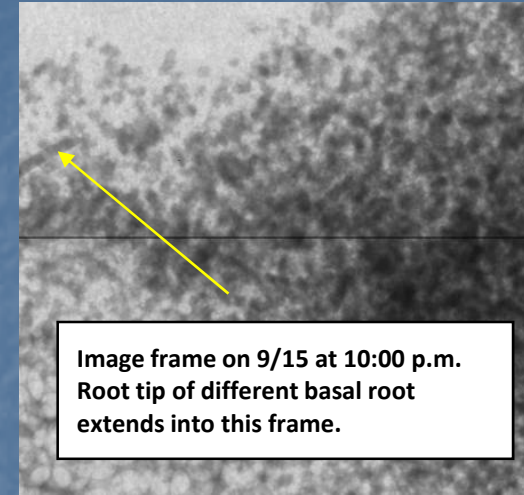




# Root Growth Occurs Day & Night



16 hr Daytime Growth



8 hr Nighttime Growth

# RootViz Application Areas

- General Root Studies
- Root Architecture
- Root Response to:
  - Nutrients
  - Water
  - Symbionts
  - Pathogens
- Phenotypic Plasticity of Root Structures
- Genetic Transformation Effects
- Functional Genomics

# Acknowledgements

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